

# Shedding Hub: An Open Science Portal for Existing Pathogen Shedding Data and Models

Yuke (Andrew) Wang, PhD, MSPH <sup>1,2,3</sup>  
yuke.wang@emory.edu

Till Hoffmann, PhD, MPhys <sup>4</sup>  
thoffmann@hsph.harvard.edu

<sup>1</sup>Hubert Department of Global Health, Rollins School of Public Health, Emory University

<sup>2</sup>Emory Center for Infectious Disease Modeling and Analytics and Training Hub, Emory University

<sup>3</sup>Center of Global Safe Water, Sanitation, and Hygiene, Emory University

<sup>4</sup>Department of Biostatistics, Harvard T.H. Chan School of Public Health, Harvard University

IDM Annual Symposium, October 1st, 2024



ROLLINS  
SCHOOL OF  
PUBLIC  
HEALTH



CGSW  
Center for Global Safe WASH

Leading and  
Learning in WASH



SCHOOL OF PUBLIC HEALTH

# Center for Infectious Disease Modeling and Analytics & Training Hub (CIDMATH)

# InsightNet (National Outbreak Analytics & Disease Modeling Network)



*Illustration of CFA's partners working to detect and control an infectious disease outbreak. <https://www.cdc.gov/insight-net/php/about/index.html>*

- Established in 2023 by CDC Center for Forecasting and Outbreak Analytics (CFA)
- Focuses on training, analytical tool development, and advancing the analysis and use of data about infectious disease spread
- brings together >100 academic and private partners and health departments

# CIDMATH



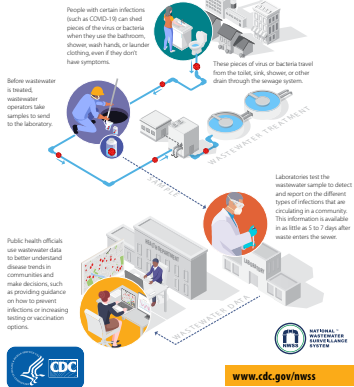
- 13 centers funded through the CFA
- Emory CIDMATH is a Center of Innovation
- Partners include Georgia DPH, Kaiser Permanente of GA, and the Georgia Emerging Infections Program (EIP)

# Motivation

# Wastewater Surveillance

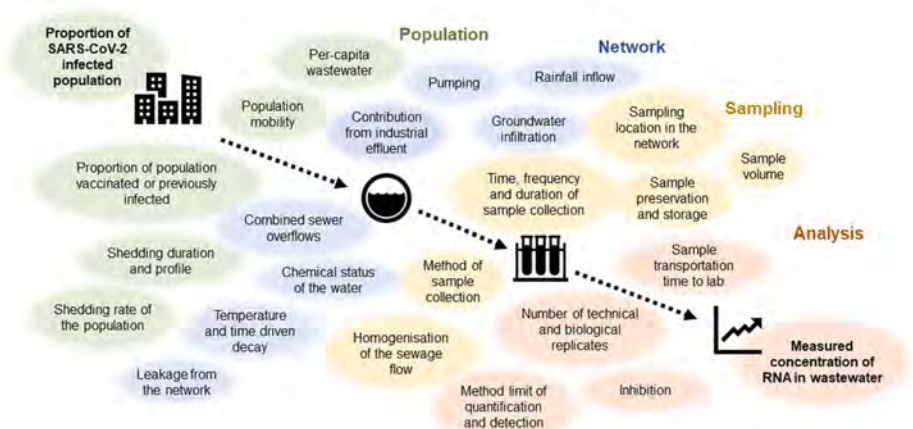
## Wastewater Monitoring HOW DOES IT WORK?

Wastewater monitoring is an early detection tool that can help communities prepare for and take action to address increasing cases of infectious diseases.

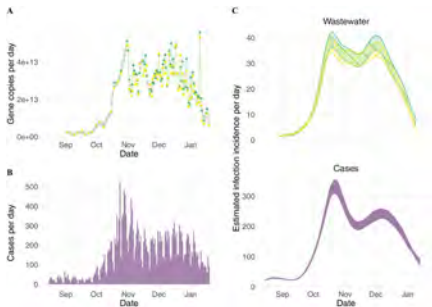


- Wastewater surveillance (WWS) is an approach for monitoring specific pathogen(s) circulating in a population by examining sewage samples.
- Pathogens shed in feces, urine, sputum, and vomit are aggregated in the sewage system.
- A well-designed WWS provides actionable information, including **certification of elimination, early warning, nowcasting/predicting trends, and identification of hotspots.**

# Sources of Uncertainty for WWS

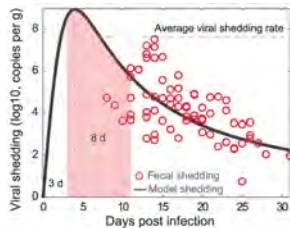


# Wastewater-based Epidemiology



Huisman et al., 2022

- The interpretation of WWS results in terms of clinical cases depends on quantitatively well-characterized shedding information for pathogens and biomarkers



Phan et al., 2023



# Current Knowledge of Shedding

- Most shedding data have been collected in clinical studies or human challenging studies
- Limited raw shedding data are openly available
- Shedding data are not standardized
- No public accessible tutorial or feasible tool for modeling shedding dynamics
- No community portal for learning and contributing shedding knowledge

# Shedding Hub

# Open Science



## Principles of Open Science:

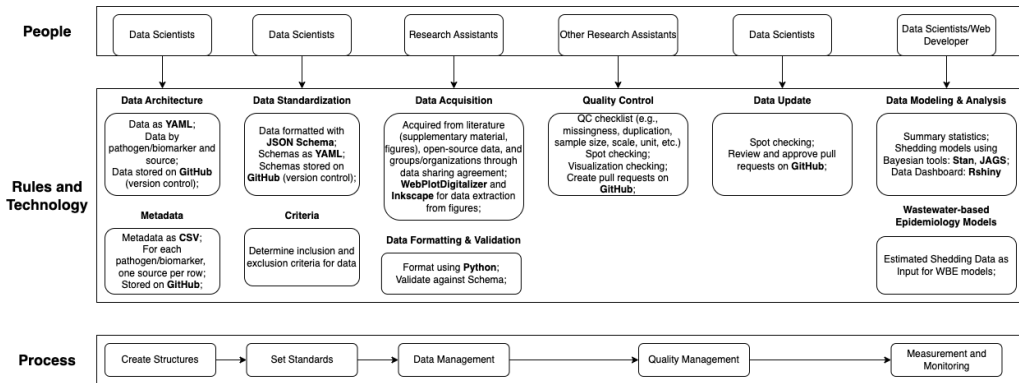
- Accessible
- Verifiable
- Reliable
- Reproducible
- Sustainable

# Shedding Hub Organization

The screenshot shows the GitHub organization page for 'shedding-hub'. At the top, there is a navigation bar with 'Overview', 'Repositories', 'Packages', 'Teams', 'People', and 'Settings'. The organization's profile includes a cartoon mascot, the name 'Shedding Hub', 2 followers, and a location in the United States of America. A 'README.md' section describes the project's goal: collating data and statistical models for biomarker shedding in human specimens. Below this, a 'Popular repositories' section lists two public repositories: 'shedding-hub' (Python) and 'shedding-hub.github.io' (HTML). On the right side, there are sections for 'View as Public', 'Discussions', and 'People'.

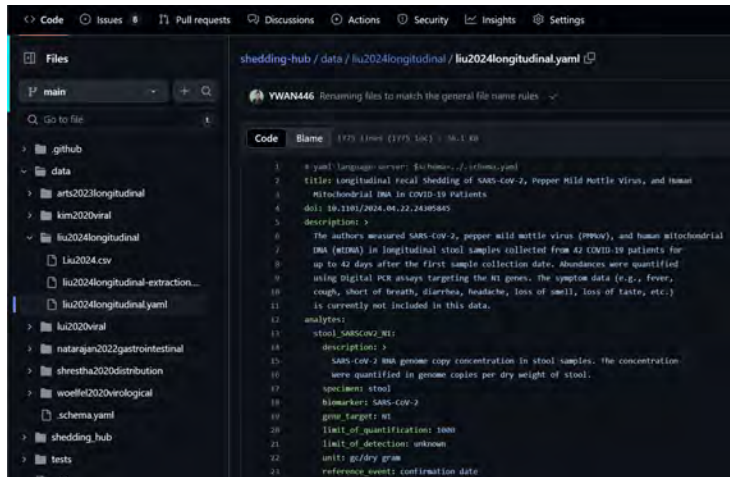
<https://github.com/shedding-hub>

# Shedding Hub Data Governance Framework



This Data Governance Framework is an extension of the [tillahoffmann/shedding](https://github.com/tillahoffmann/shedding) repository.

# Data Structure

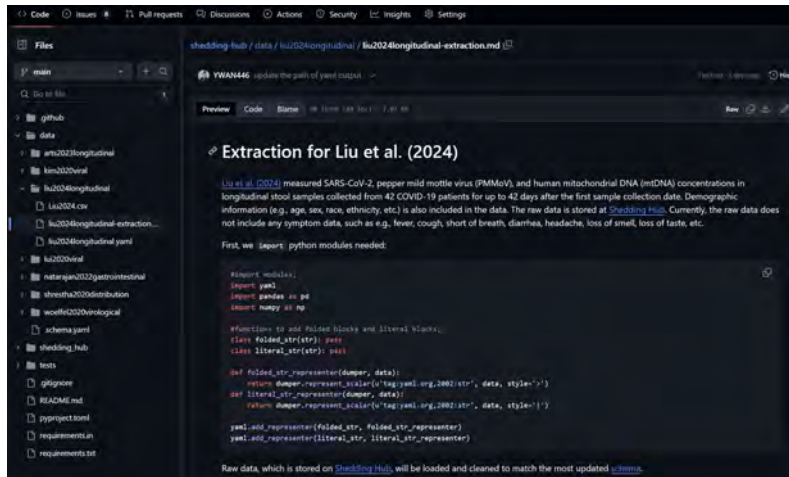


The screenshot shows a GitHub repository interface. The left sidebar displays the file structure under the 'main' branch, with the 'data' directory expanded to show 'liu2024longitudinal'. The main content area shows the 'liu2024longitudinal.yaml' file, which is a YAML file describing a dataset. The file content is as follows:

```
1 # yaml language server: $(dirname $0)/.ydlms.yaml
2 title: longitudinal fecal shedding of SARS-CoV-2, Pepper Mild Mottle Virus, and Human
3 Mitochondrial DNA in COVID-19 Patients
4 doi: 10.1101/2024.04.22.24105845
5 description: >
6 The authors measured SARS-CoV-2, pepper mild mottle virus (PMMoV), and human mitochondrial
7 DNA (mtDNA) in longitudinal stool samples collected from 42 COVID-19 patients for
8 up to 42 days after the first sample collection date. Abundances were quantified
9 using Digital PCR assays targeting the N1 genes. The symptom data (e.g., fever,
10 cough, short of breath, diarrhea, headache, loss of smell, loss of taste, etc.)
11 is currently not included in this data.
12 analytes:
13 stool_SARSCoV2_N1:
14   description: >
15     SARS-CoV-2 RNA genome copy concentration in stool samples. The concentration
16     were quantified in genome copies per dry weight of stool.
17   specimen: stool
18   biomarker: SARS-CoV-2
19   gene_target: N1
20   limit_of_quantification: 1000
21   limit_of_detection: unknown
22   unit: gc/dry gram
23   reference_event: confirmation date
```

- Includes raw data files, markdown files to process data, standardized data in YAML format, and a schema file
- Public accessible on GitHub

# Data Processing



```
shedding-hub / data / liu2024longitudinal / liu2024longitudinal-extraction.md
YWAN446 update the path of yaml output
Preview Code Blame 18 lines 188 loc 1 7.4k 46
Raw

Extraction for Liu et al. (2024)

Liu et al. (2024) measured SARS-CoV-2, pepper mild mottle virus (PMMoV), and human mitochondrial DNA (mtDNA) concentrations in longitudinal stool samples collected from 42 COVID-19 patients for up to 42 days after the first sample collection date. Demographic information (e.g. age, sex, race, ethnicity, etc.) is also included in the data. The raw data is stored at Shedding Hub. Currently, the raw data does not include any symptom data, such as e.g., fever, cough, short of breath, diarrhea, headache, loss of smell, loss of taste, etc.

First, we report python modules needed:

!important modules:
!important yaml
!important pandas as pd
!important numpy as np

#Functions to add folded blocks and literal blocks:
class folded_str(str): pass
class literal_str(str): pass

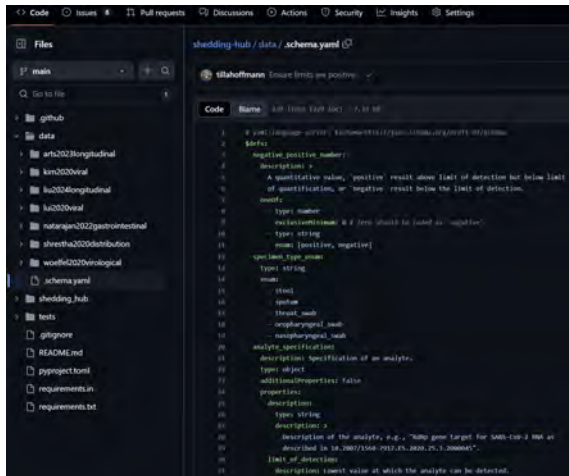
def folded_str_representer(dumper, data):
    return dumper.represent_scalar(u'tag:yaml.org,2002:str', data, style='')
def literal_str_representer(dumper, data):
    return dumper.represent_scalar(u'tag:yaml.org,2002:str', data, style='|')

yaml.add_representer(folded_str, folded_str_representer)
yaml.add_representer(literal_str, literal_str_representer)

Raw data, which is stored on Shedding Hub, will be loaded and cleaned to match the most updated schema.
```

- **Verifiable** with information sources
- **Reproducible** with open code

# Data Standardization



```
shedding-hub / data / .schema.yaml
silahoffmann Ensure limits are positive ✓

Code Blame 3.0K 1.00K 1.2K 20K 7, 18 18

1 # yaml-language-server: $schema=https://json-schema.org/draft/2019-09/schema
2 $defs:
3   negative_positive_number:
4     description: >
5       A quantitative value, 'positive' result above limit of detection but below limit
6       of quantification, or 'negative' result below the limit of detection.
7     minOf:
8       - type: number
9       - exclusiveMinimum: 0.0 (zero should be used as 'negative')
10      - type: string
11      min: [positive, negative]
12
13   spec_item_type_union:
14     type: string
15     minOf:
16       - stool
17       - spatula
18       - throat_swab
19       - oropharyngeal_swab
20       - nasopharyngeal_swab
21
22   analyte_specification:
23     description: Specification of an analyte.
24     type: object
25     additionalProperties: false
26     properties:
27       description:
28         type: string
29         description: >
30           Description of the analyte, e.g., "hmp gene target for SARS-CoV-2 RNA as
31           described in 10.1093/infdis/jiaa25.1.2000045".
32
33     limit_of_detection:
34       description: Lowest value at which the analyte can be detected.
```

- JSON schema ([json-schema.org](https://json-schema.org)) is a standard to specify the structure of data, validate it, and include documentation about each field
- All data uploaded will be validated against the schema automatically



# Data Validation

The screenshot shows a GitHub pull request titled "Create shrestha2020distribution.yaml #35". The pull request is merged and includes a comment from WenfeiXiao. The comment discusses a study on COVID-19 transmission, mentioning 528 tests at the Cleveland Clinic and a discrepancy in the limit of detection (LOD) between a paper (20 pg/mL) and a dataset (67.24 pg/mL). The pull request also includes a table of statistics for the data.

WenfeiXiao commented last week

This study evaluated the transmission potential of COVID-19 by examining viral load over time. Over six weeks, 230 healthcare personnel underwent 528 tests at the Cleveland Clinic. Cycle threshold (Ct) values were obtained using RT-PCR targeting the N gene, and viral loads were calculated. Data were obtained from the combined dataset in the supplementary materials of Challenger et al. BMC Medicine (2022) 20:25 (<https://doi.org/10.1186/s12916-021-02220-0>).

There is a discrepancy between the paper suggesting limit\_of\_detection as 20 pg/mL and Challenger combined dataset suggesting limit\_of\_detection as 67.24 pg/mL.

github-actions bot committed last week • edited •

Summary for changed file: data/shrestha2020distribution.yaml:

	n_samples	n_unique_participants	n_negative	n_positive	n_quantified	min	max
mask_SARS-CoV-2	528	230	119	8	89	436.779904	1.809101-679416

To generate **reliable** data:

- Data will be checked and reviewed by at least two reviewers
- Functions were created to generate automatic data summary
- Conversations and decisions will be documented on GitHub

# Modeling Tutorial and Code

- Introduction
- Exponential Decay Model
- Gamma Model
- References

## Bayesian Workflow for Modeling Shedding Dynamics using Rstan

Yuke Wang, Hubert Department of Global Health, Rollins School of Public Health, Emory University  
 Till Hoffmann, Department of Biostatistics, Harvard T.H. Chan School of Public Health, Harvard University  
 2024-09-13

### Introduction

The current tutorial demonstrates the Bayesian workflow to model shedding data using rstan. We use the longitudinal SARS-CoV-2 fecal shedding data from Miller et al. (2021). The data includes observations of SARS-CoV-2 RNA concentration in 82 stool samples from 8 patients. The date of sample collection spans from Day 3 to Day 22 after symptom onset. We load the data in R with:

```
shedding_data <- stana::read(subjects = 421, 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82)

## # 82x3 matrix of RNA concentration (copies/ml)
## #      0 1 2
## # 1 0.000 0.000 0.000
## # 2 0.000 0.000 0.000
## # 3 0.000 0.000 0.000
## # 4 0.000 0.000 0.000
## # 5 0.000 0.000 0.000
## # 6 0.000 0.000 0.000
## # 7 0.000 0.000 0.000
## # 8 0.000 0.000 0.000
## # 9 0.000 0.000 0.000
## # 10 0.000 0.000 0.000
## # 11 0.000 0.000 0.000
## # 12 0.000 0.000 0.000
## # 13 0.000 0.000 0.000
## # 14 0.000 0.000 0.000
## # 15 0.000 0.000 0.000
## # 16 0.000 0.000 0.000
## # 17 0.000 0.000 0.000
## # 18 0.000 0.000 0.000
## # 19 0.000 0.000 0.000
## # 20 0.000 0.000 0.000
## # 21 0.000 0.000 0.000
## # 22 0.000 0.000 0.000
## # 23 0.000 0.000 0.000
## # 24 0.000 0.000 0.000
## # 25 0.000 0.000 0.000
## # 26 0.000 0.000 0.000
## # 27 0.000 0.000 0.000
## # 28 0.000 0.000 0.000
## # 29 0.000 0.000 0.000
## # 30 0.000 0.000 0.000
## # 31 0.000 0.000 0.000
## # 32 0.000 0.000 0.000
## # 33 0.000 0.000 0.000
## # 34 0.000 0.000 0.000
## # 35 0.000 0.000 0.000
## # 36 0.000 0.000 0.000
## # 37 0.000 0.000 0.000
## # 38 0.000 0.000 0.000
## # 39 0.000 0.000 0.000
## # 40 0.000 0.000 0.000
## # 41 0.000 0.000 0.000
## # 42 0.000 0.000 0.000
## # 43 0.000 0.000 0.000
## # 44 0.000 0.000 0.000
## # 45 0.000 0.000 0.000
## # 46 0.000 0.000 0.000
## # 47 0.000 0.000 0.000
## # 48 0.000 0.000 0.000
## # 49 0.000 0.000 0.000
## # 50 0.000 0.000 0.000
## # 51 0.000 0.000 0.000
## # 52 0.000 0.000 0.000
## # 53 0.000 0.000 0.000
## # 54 0.000 0.000 0.000
## # 55 0.000 0.000 0.000
## # 56 0.000 0.000 0.000
## # 57 0.000 0.000 0.000
## # 58 0.000 0.000 0.000
## # 59 0.000 0.000 0.000
## # 60 0.000 0.000 0.000
## # 61 0.000 0.000 0.000
## # 62 0.000 0.000 0.000
## # 63 0.000 0.000 0.000
## # 64 0.000 0.000 0.000
## # 65 0.000 0.000 0.000
## # 66 0.000 0.000 0.000
## # 67 0.000 0.000 0.000
## # 68 0.000 0.000 0.000
## # 69 0.000 0.000 0.000
## # 70 0.000 0.000 0.000
## # 71 0.000 0.000 0.000
## # 72 0.000 0.000 0.000
## # 73 0.000 0.000 0.000
## # 74 0.000 0.000 0.000
## # 75 0.000 0.000 0.000
## # 76 0.000 0.000 0.000
## # 77 0.000 0.000 0.000
## # 78 0.000 0.000 0.000
## # 79 0.000 0.000 0.000
## # 80 0.000 0.000 0.000
## # 81 0.000 0.000 0.000
## # 82 0.000 0.000 0.000
```

As most samples collected in shedding studies are several days after symptom onset when the number of pathogens shed is decreasing, in this tutorial, we focus on modeling the decay phase of the shedding dynamics. We consider two classical models, **exponential decay model** and **gamma model**.

### Exponential Decay Model

When the concentration of pathogen shed  $c(t)$  is subject to **exponential decay**, it decreases at a rate  $\lambda_0$  proportional to the current value:

$$c(t) = c_0 e^{-\lambda_0 t}$$

Where  $c_0$  is the concentration of pathogen shed at symptom onset and  $t$  is the day after symptom onset.

When there are samples from multiple subjects, we can develop a hierarchical model:

$$c_i(t) = c_{i0} \mu^{-\lambda_0 t}$$

for subject  $i$  ( $\log(\mu_{i0}), \log(\lambda_{i0}) \sim N(\mu, \Sigma^{-1})$ , where  $\mu = (\mu_{i0}, \lambda_{i0})$ ). And we use the Cholesky decomposition for the correlation matrix and  $\Sigma^{-1} = \text{diag}(\tau_0, \tau_1) L L^T \text{diag}(\tau_0, \tau_1)$ .  $L$  is lower triangular such that  $L L^T$  is positive definite.

```
without_rstan <- "without_rstan.R"
library(rstan)
library(rstanarm)
library(cowplot)
library(ggplot2)
library(dplyr)
library(tibble)
library(purrr)
library(reshape2)
```

### Single Subject without Censored Data

First, we consider the simplest case of one subject with only quantifiable positive samples. For this example, we select Subject 1.

- Tutorial for modeling shedding data in Python/R
- **Reproducible** using markdown files

# Contribution

## Shedding Hub Shedding Hub

The Shedding Hub collates data and statistical models for biomarker shedding (such as viral RNA or drug metabolites) in different human specimen (such as stool or sputum samples). Developing wastewater-based epidemiology into a quantitative, reliable epidemiological monitoring tool motivates the project.

Datasets are extracted from appendices, figures, and supplementary materials of peer-reviewed studies. Each dataset is stored as a [.yaml](#) file and validated against our [data schema](#) to verify its integrity.

### Contributing

Thank you for contributing your data to the Shedding Hub and supporting wastewater-based epidemiology! If you hit a bump along the road, [create a new issue](#) and we'll sort it out together.

We use [pull requests](#) to add and update data, allowing for review and quality assurance. Learn more about the general workflow [here](#). To contribute your data, follow these easy steps (if you're already familiar with pull requests, steps 2 and 3 are for you):

1. Create a [fork](#) of the Shedding Hub repository by clicking [here](#) and [clone](#) the fork to your computer. You only have to do this once.
2. Create a new `my_cool_study/my_cool_study.yaml` file in the `data` directory and populate it with your data. See [here](#) for a comprehensive example from [Wobbel et al. \(2020\)](#). A minimal example for studies with a single analyte (e.g., SARS-CoV-2 RNA concentration in stool samples) is available [here](#), and a minimal example for studies with multiple analytes (e.g., crAssphage RNA concentration in stool samples and caffeine metabolites in urine) is available [here](#).
3. Optionally, if you have a recent version of [Python](#) installed, you can validate your data to ensure it has the right structure before contributing it to the Shedding Hub.
  - Run `pip install -r requirements.txt` from the command line to install all the Python packages you need.
  - Run `pytest` from the command line to validate all datasets, including the one you just created.
4. Create a new [branch](#) by running `git checkout -b my_cool_study`. Branches let you isolate changes you are making to the data, e.g., if you're simultaneously working on adding multiple studies—much appreciated! You should create a new branch from the `main` branch for each dataset you contribute; see [here](#) for more information.
5. Add your changes by running `git add data/my_cool_study/my_cool_study.yaml` and commit them by running `git commit -m "Add data from Someone at al. (20xx)."'`. Feel free to pick another commit message if you prefer.

- **Sustainable** with partnerships and contributions from the research community

# Shedding Hub Website

Shedding Hub

10,529 biomarker measurements for 455 participants from 7 studies. And counting.

**Longitudinal and quantitative fecal shedding dynamics of SARS-CoV-2, pepper mild mottle virus, and crAssphage**

Participants: 48

Measurements: 1,502

Biomarkers: SARS-CoV-2, PMMoV, crAssphage

The authors present longitudinal, quantitative fecal shedding data for SARS-CoV-2 RNA, pepper mild mottle virus (PMMoV) RNA, and crAss-like phage (crAssphage) DNA from 48 COVID-19 patients. Abundances were quantified using (RT)-ddPCR assays targeting the N and ORF1a genes. The data were obtained from supplementary material.

[View Source](#) [View on GitHub](#) [Explore Dataset](#)

Shedding Hub Data and statistical models for biomarker shedding

**Longitudinal and quantitative fecal shedding dynamics of SARS-CoV-2, pepper mild mottle virus, and crAssphage**

The authors present longitudinal, quantitative fecal shedding data for SARS-CoV-2 RNA, pepper mild mottle virus (PMMoV) RNA, and crAss-like phage (crAssphage) DNA from 48 COVID-19 patients. Abundances were quantified using (RT)-ddPCR assays targeting the N and ORF1a genes. The data were obtained from supplementary material.

**Analytes**

**ctac1\_SARScov2\_N**

Concentration of RNA of the N gene quantified using (RT)-ddPCR in stool samples. The concentration was quantified in gene copies per dry weight of stool. The limit of blank (LOB), determined as the upper 95% confidence limit of the negative extraction control, ranged from 11.2 to 1,550 gc/dry weight. The reported number is either the measured concentration of SARS-CoV-2 N or the LOB if the concentration wasn't detectable.

- Biomarker: SARS-CoV-2
- Specimen: stool
- Units: gc/dry gram
- Gene target: N
- Participants: 48
- Negative samples: 184
- Positive samples (not quantifiable): 0
- Quantifiable samples: 193
- Limit of quantification: unknown
- Limit of detection: unknown
- Limit of blank: 1,550.00

**ctac1\_SARScov2\_ORF1a**

Concentration of RNA of the ORF1a gene quantified using (RT)-ddPCR in stool samples. The concentration was quantified in gene copies per dry weight of stool. The limit of blank (LOB), determined as the upper 95% confidence limit of the negative extraction control, ranged from 11.2 to 1,550 gc/dry weight. The reported number is either the measured concentration of SARS-CoV-2 ORF1a or the LOB if the concentration wasn't detectable.

- Biomarker: SARS-CoV-2
- Specimen: stool
- Units: gc/dry gram
- Gene target: ORF1a
- Participants: 48
- Negative samples: 186
- Positive samples (not quantifiable): 0
- Quantifiable samples: 191
- Limit of quantification: unknown
- Limit of detection: unknown
- Limit of blank: 1,550.00

shedding-hub.github.io

# Work Plan

<i>Phase 1</i>	<i>Phase 2</i>	<i>Phase 3</i>
<b>Objectives:</b> Build a Shedding Hub Team; Develop the Basic Structure and Workflow	<b>Objectives:</b> Expand to Additional Pathogens and Biomarkers; Invite Contribution of Data and Models	<b>Objectives:</b> Develop analytical tools (e.g., dashboard, packages) and promote usage and contribution
<b>Data Sources:</b> Open Access Data, Published in Literature <b>Priority:</b> Quantifiable Longitudinal Measurements	<b>Data Sources:</b> Limited Access Data <b>Priority:</b> Quantifiable Longitudinal Measurements	<b>Priority:</b> Semi-quantifiable (CT values) or Non-quantifiable (Presence/Absence) Measurements
<b>Models:</b> Developed within the Shedding Hub Team	<b>Models:</b> Published Models by Experts	<b>Models:</b> Contributed Models by Research Community
<b>Prioritized Biomakers:</b> Pathogens of Interest for WWS	<b>Prioritized Biomakers:</b> Additional Pathogens and Biomarkers	<b>Prioritized Biomakers:</b> Additional Pathogens and Biomarkers
<i>May 2024 – Sep 2025</i>	<i>Oct 2025 – Sep 2026</i>	<i>Oct 2026 – Sep 2027</i>

# Use Cases

- To support wastewater-based epidemiology applications estimating disease incidence for various infectious diseases
- To better understand of sensitivity of different disease diagnostic methods (nasopharyngeal swab vs. rectal swab)
- To support decision making for disease control and prevention policys (e.g., how long the quanrantine period should be?)
- To support wastewater monitoring for drug use

# Acknowledgements

## Shedding Hub Team



Yuke Wang



Till Hoffmann



Weifei Xiao



Youwei Hu



Zirui Chen



Haisu Zhang

## CIDMATH Team

